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STUDY THE BIOLOGICAL EFFECT AND NANO PREPARATION OF OLEANDER PLANT *NERIUM OLEANDER* ON THE HAIRY GRAIN BEETLE *Trogoderma granarium* EVERTS (COLEOPTERA: DERMESTIDAE)

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ABSTRACT

Cereal crops, including wheat (*Triticum aestivum*), are among the most important vital and strategic crops in the world because they contain important food sources of vitamins, proteins, carbohydrates, fats, and other nutrients. Stored grains are affected by many insect pests, causing significant economic losses in grain weight that may reach 10% in one storage season. One of the most dangerous stored insects that infest grains is the hairy grain beetle (*Trogoderma granarium*).

The insect is known for its resistance to various liquid and gaseous chemical pesticides, as well as its ability to withstand long periods of drought and high temperatures. This adaptability has facilitated its spread across different countries and enabled it to conquer new geographical areas. Several methods have been employed to combat this insect, with chemical control being the most prominent. However, the use of chemical pesticides has resulted in negative environmental impacts and challenges in addressing the various roles insects play.

Due to the economic significance of the hairy grain beetle and the need to minimize reliance on harmful chemical pesticides, plant extracts and nano-preparations of iron from the leaves of the oleander plant (*Nerium oleander*) were utilized. These compounds have shown high effectiveness in controlling the Khabra insect in both laboratory and storage environments, while also being environmentally safe.

Keywords: *Nerium oleander*, proteins, pesticides, insects.

دراسة التأثير البيولوجي والمستحضر النانوي لنبات الدفلة على خنفساء الحبوب الشعيرية *Trogoderma granarium* (Coleoptera : Dermestidae) Everts

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الخلاصة

تعد محاصيل الحبوب ومنها محصول الحنطة (*Triticum aestivum*) من اهم المحاصيل الحيوية والاستراتيجية في العالم بسبب احتوائها على مصادر غذائية مهمة من فيتامينات وبروتينات وكربوهيدرات ودهون وعناصر غذائية أخرى. تتأثر الحبوب المخزونة بالعديد من الآفات الحشرية مسببة خسائر اقتصادية كبيرة في وزن الحبوب قد تصل الى 10% في موسم خزني واحد. ومن اخطر الحشرات المخزنية المصنفة عالميا التي تصيب الحبوب المخزونة حشرة خنفساء الحبوب الشعيرية *Trogoderma granarium*.

وتعرف الحشرة بمقاومتها لمختلف المبيدات الكيماوية السائلة والغازية، فضلا عن قدرتها على تحمل فترات طويلة من الجفاف ودرجات الحرارة المرتفعة. وقد سهلت هذه القدرة على التكيف انتشاره عبر مختلف البلدان ومكنته من غزو مناطق جغرافية جديدة. وقد تم استخدام طرق متنوعة لمكافحة هذه الحشرة، وكان أبرزها مكافحة الكيماوية. إلا أن استخدام المبيدات الكيماوية أدى إلى آثار بيئية سلبية وتحديات في معالجة ادوار الحشرة المختلفة. نظراً للأهمية الاقتصادية لخنفساء الحبوب الشعيرية ولتقليل الاعتماد على المبيدات الكيماوية الضارة، تم استخدام المستخلصات النباتية والمستحضرات النانوية للحديد المستخلص من أوراق نبات الدفلة (*Nerium oleander*) حيث أثبتت هذه المركبات فعالية كبيرة في مكافحة حشرة الخابرا مختبريا ومخزنيا، كما أنها آمنة بيئيا.

الكلمات المفتاحية: الدفلة، بروتين، مبيدات، حشرات.

INTRODUCTION

Wheat (*T. aestivum*) is a strategic crop that is important for food security for the world's population, and its global demand is expected to reach 3.3 billion tons by 2050 (Mustafa, 2017; UN report, 2022). Stored insects are among the most important pests that infect wheat seeds in stores, damaging them and reducing their quality, causing significant economic losses in the quality and weight of grains estimated at 6-73% (Khalique et al., 2018; Rajendran, 2020). The most important of these is the hairy grain beetle (Khabra) *T. granarium* (Everts), which is one of the dangerous and globally widespread insects and is characterized by its voracious feeding on completely stored grains in addition to its waste that is harmful to health (Khan et al., 2019; Athanassiou et al., 2019).

The widespread and frequent use of chemical pesticides to control stored insects, especially the grain beetle, has led to the emergence of resistance to pesticides, in addition to environmental pollution and toxicity to humans and animals. The most used materials are Phostoxin tablets and strips that produce phosphine gas (Wakil et al., 2021; Yılmaz & Koçak, 2022). To reduce the risks of using pesticides, alternative methods have begun to be used for chemical pesticides that are safe for the environment and human health, such as the use of plant extracts that contain many biological compounds that are effective in controlling pests and



protecting stored grains from insect infestation, which are environmentally friendly compounds due to their biodegradability and low toxicity to animals (Alvi *et al.*, 2018).

The use of nano-preparations is one of the most modern and effective methods for combating stored pests. Plant extracts are used in the biosynthesis of nanoparticles and improving the control properties. It is an environmentally safe and economical method due to the speed and ease of preparation. It is expected that the global production of nano-preparations will reach ten thousand tons during the year 2026 (Ovais *et al.*, 2018).

The economic importance of the grain beetle and its damages It belongs to the order of coleoptera, its scientific name is *T. granarium*, and it is considered one of the most important insect pests that attack stored grains and lead to their spoilage and the most dangerous in many countries of the world and in Iraq, causing a large economic loss in stored grains in terms of quantity and quality. In addition to the secretion of molting skins, dead insects, and insect waste, causing grain contamination, and as a result of the insect feeding in large numbers due to its rapid reproduction, it leads to an increase in the temperature of grain stores, which exposes them to infection with fungi and bacteria and makes them unfit for marketing, human consumption, and use as seeds (Media, 2007; Ahmedani *et al.*, 2009).

The harmful stage of the insect is the larval stage, especially in advanced ages. The larvae feed on grains by gnawing them and destroying the embryo and endosperm, which makes them unsuitable for cultivation due to the low germination rate. The insect is characterized by its voracious feeding on different types of grains and dry food. One larva can consume 18 mg of grains per day, leaving behind waste at a rate of 14.8 mg. It is characterized by activity in a wide temperature range between 20- and 50°C, and the loss rate reaches 70% when suitable conditions are available (Al-Iraqi, 2010).

The importance of oleander plant extracts in pest controls. The ethanolic extract of *N. oleander* flowers has proven highly effective against the activity of plant pathogenic fungi, including *Fusarium oxysporum*, *Fusarium solani*, *Alternaria alternate*, and *Rhizoctonia solani* (Hadizadeh *et al.*, 2009). The bark, stem, leaves, flowers, and roots extract of oleander have also been used as an anti-feeding insecticide to control the larvae of the diamondback moth *Plutella xylostella* and *Aedes aegypti* (Komalamisra *et al.*, 2005). The hexane and aqueous extracts of crude oleander flowers were highly effective in mortality of *Culex quinquefasciatus* larvae, with LC50 values reaching 102.54 and 61.11 ppm after 24 and 48 hours of treatment with the hexane extract (Raveen *et al.*, 2014).

Al-Iraqi (2003) studied the efficiency of several plant powders against the hairy grain beetle (Khabra) *T. granarium*, including *N. oleander* L. powder, which was mixed with wheat grains at concentrations of 0.3, 0.6, and 0.9%. It influenced the generation duration, reaching 48.83, 48.96, and 54.09 days for the concentrations, respectively. It reached 40.18 days in the control treatment due to the anti-feeding effect of the oleander plant, as it reduced egg laying. Suleiman (2005) found that surface treatment of the adults of the rusty flour beetle *Tribolium castaneum* (Herbst.) with alcoholic and aqueous extracts of *N. oleander* L. leaves caused mortality of adults after 48 hours of treatment.



The cold aqueous extract of *N. oleander* leaves influenced the periodic death of eggs and larvae of the greater wax moth *Galleria mellonella* at several concentrations, as the mortality rate at the concentration of 4 mg/ml reached (99.4% and 96.6%) for eggs and first-stage larvae, respectively (Najib, 2019). Al-Mashhadani (2012) also found that the powder of *N. oleander* leaves had a high repellent rate against larvae and adults of the hairy grain beetle *T. granarium* in the test he conducted to test the attraction and repellent of the powder. A study showed that the extract of *N. oleander* L. caused an inhibition in the severity of the disease early blight on tomato crop caused by the fungus *Alternaria solani* (Nashwa & Abu-Elyousr, 2012).

Nanotechnology

The word nano refers to nanoparticles that are between 1 and 100 nanometers in size. Nanotechnology has spread rapidly in the world due to the features and properties of nanoparticles, their wide applications, their chemical composition, and their surface structure. Their small size increases the surface cohesion of nanoparticles (Adhikari et al., 2013; Tawfeeq, 2014; Ahmida et al., 2017; Gulzar et al., 2020). Nanoparticles that have been converted into pesticides are characterized by green technology that selectively targets insects to preserve plants and the environment, as well as combating insects that consume stored grains (Sahayaraj, 2017).

The importance of nanoparticles in insect control:

The extract of the oleander plant was used to prepare silver nanoparticles by reducing silver nitrate and studied its effect in preventing *Culex pipiens mosquito* larvae from reaching the adult stage by using several concentrations of the plant extract and silver nanoparticles. The silver nanoparticles were more effective in preventing the emergence of adult insects from the extract by about 33.33 times, and both the plant extract and silver particles led to deformities in the treated larvae, which led to mortality of the insect and failure to complete its life cycle (Al-Qurashi et al., 2015). (Rouhani et al., 2008) showed that treating the food of the southern cowpea beetle *Callosobruchus maculatus* with silver nanoparticles at a concentration of 2.5 g/kg grains led to their death by 75% after 14 days of treatment. Silver nanoparticles prepared using the aqueous extract of *Euphorbia prostste* leaves were used to control the adults of the rice weevil *Sitophilus oryzae* at a concentration of 250 mg/kg and a size of 25–80 nm, and the mortality rate reached 100% after 14 days of treatment (Zahir et al., 2012).

(Chakravarthy et al. 2012) found that nanoparticles of titanium oxide 2TiO, silver AgNPS and cadmium sulphate Cds at a concentration of 2400 ppm used to control the cotton leafworm *Spodoptera lituralis* had a mortality rate of (73, 56 and 93%), respectively. (Al-Shammari, 2015) showed that silver nanoparticles prepared with aqueous extract of eucalyptus leaves at a concentration of 4250 ppm were used to control mealybugs, with a mortality rate of adults reaching 66.7% within 72 hours of treatment. (Al-Shujairi, 2018) prepared silver nanoparticles using alcoholic extract of myrtle leaves and used them against nymphs and adults of cotton *A. gossypii*, with a mortality rate of 99.2% after 72 hours of treatment.



(Najib, 2019) found that silver nanoparticles prepared from the cold aqueous extract of *N. oleander* and *Ricinus communis* leaves had an effect in inhibiting the eggs and mortality the larvae of the greater wax moth *G. mellonella*, as the percentage of egg-stage inhibiting at a concentration of 4 mg/ml reached 98.56% and 100% for the two plants, respectively, and the percentage of mortality the first-stage larvae reached 92.3% and 100%, respectively, at the same concentration.

The importance of nanoparticles in controlling the grain beetle *T. granarium*:

Al-Hayali (2018) proved that treating wheat grains with nanoparticles EoEC-AAgNPs prepared from *Eucalyptus camaldulensis* oil and *Artemisia herba alba* at a concentration of 8000 ppm influenced mortality in the larvae of the grain beetle *T. granarium* with a mortality rate of 96.66% after 48 hours of treatment. The nanoparticles Agnps-Eoec prepared from *Artemisia herba alba* extract and *E. camaldulensis* oil, which were mixed with wheat grains at a concentration of 8000 ppm, moralized the larvae of the grain beetle *T. granarium* with a mortality rate of 96.66% within 48 hours (Al-Hayali, 2018). (Almadiy et al. 2018) found that the use of silver nanoparticles prepared using the alcoholic extract of Peganmum harmala seeds was highly effective in killing larvae and adults of the hairy grain beetle *T. granarium*.

Uses of iron nanoparticles in insect control:

Iron nanoparticles are an environmentally friendly magnetic nanomaterial used as a pesticide against the larvae of the Egyptian mosquito *Aedes aegypti*. Its nanomaterial consists of two materials (SAMNs&chlorin), which are iron oxide nanoparticles γ -Fe₂O₃ and are called surface active nanoparticles SAMNs representing the core (Surface Active Maghemite Nanoparticles SAMNs) and chlorin that forms the shell (chlorin-e6). It has a toxic effect on the larvae, and the material is characterized by its chemical effectiveness and stability and is a safe alternative to synthetic insecticides and has no harm to the environment (Magro et al., 2019). (Chen et al., 2014) studied the toxic effect of iron nanoparticles on the fruit fly *Drosophila melanogaster*. Reda et al. (2016) conducted a study to evaluate the effectiveness of iron oxide nanoparticles (α -Fe₂O₃) against the larvae of the mosquito *C. pipiens*. The particles were prepared chemically using low temperature and had a lethal effect on larvae in the laboratory at concentrations of 0.025, 0.25, 2.5, and 25 ppm of the nanomaterial, as the larval mortality rates reached 6.67, 23.34, 35, and 68.34% for the four concentrations, respectively, after 48 hours of treatment.

Iron nanoparticles were used as an alternative to chemical pesticides to study their effectiveness against the larvae and pupae of the mosquito *Culex quinquefasciatus*, Iron nanoparticles Fe₀ were prepared using the aqueous extract of *Ficus natalensis*, and iron oxide nanoparticles (Fe₂O₃) were prepared chemically. Both nanomaterials proved their efficiency in mortality the larvae and pupae of mosquitoes (Murugan et al., 2018). (Zhou et al., 2012) conducted a study to evaluate the toxicity of iron oxide nanoparticles against the discoid-cockroach *Blaberus discoidalis* by injecting iron nanoparticles into the central nervous system



of the insect. The results showed a decrease in the activity of the treated insects, which proves that iron oxide nanoparticles caused neurotoxicity to the insects, which led to a decrease in their survival rate after 40 days of injection.

Iron nanoparticles (magnetite Fe₃O₄) showed high toxicity against fruit fly *D. melanogaster* by reducing their lifespan by 30% at low concentrations (Vecchio et al., 2013). (Vega-Alvarez et al. 2014) confirmed that iron oxide nanoparticles caused the highest level of lethality in *Drosophila embryos* when compared with 8 other types of nanoparticles. A study was conducted on the effectiveness of nano-iron prepared with the alcoholic extract of *Dodonaea viscosa* L. leaves in influencing some aspects of the insect's life and its effect on the lethality of the stages of the greater wax moth *G. mellonella* L., using concentrations of 2, 5, and 10%. The direct treatment of the insect stages and the indirect treatment of the larval food proved the effectiveness of nano-iron particles compared to the alcoholic extract of *Dodonaea* leaves, as the percentage of inhibition of hatching of eggs treated superficially and at the age of 24 hours increased to 87.89, 91.13, and 93.34%, respectively, for the nano-preparation, while it reached 78.95, 85.86, and 87.04% for the concentrations, respectively, for the alcoholic extract, and it had a significant effect on some of the insect's life phenomena (Al-Jubouri, 2020).

CONCLUSION

Extracts and nano-preparations from *Diphla* plant sources can be used in the field of insect control in general and store insects in particular after recording a significant effect in lethality insects directly or indirectly (Sahi, 2019) and can be used in the field of integrated control.

REFERENCE

1. Adhikari, A.; Ghoshand, U. & Chandra, G. (2013). Nanoparticles of herbal origin: A recent eco – friend friend in mosquito control. *Asian pacific Journal Tropical Disease*. 3(2):167-168.
2. Ahmedani, M. S.; Haque, M. I.; Afzal, S. N.; Aslam, M. & Naz, S. (2009). Varietal changes in nutritional composition of wheat kernel (*Triticum aestivum* L.) caused by Khapra beetle infestation. *Pakistan Journal Botany*. 41(3) :1511-1519.
3. Ahmida, M. H.; Hamza N. A. & Aziza A. A. (2017). Simplified introduction to nanotechnology. Define its terminology, applications and presence in the environment. *Journal of the Libyan International Medical University*. 2 (1): 12- 26.
4. Al-Hayali, T. S. A. (2018). *Effect of Eucalyptus camaldulensis and Artemisia herba alba essential oils and nanomaterials and powders in the fight against the capillary beetle Trogoderma granarium* Evert (Coleoptera: Dermestidae). Ph.D. Thesis. Faculty of Agriculture. University of Baghdad. 140.
5. Al-Iraqi, R. A. (2003). Effect of powders of some plants on the capillary grain beetle (Coleoptera: Dermestidae) *Trogoderma granarium* Everts. *Arab Journal of Plant Protection*. 101-96:21.

6. Al-Iraqi, R. A. (2010). *Pests of cereals and stored materials and methods of combating them*. Dar Ibn al-Atheer for printing and publishing. University of Mosul. 616 p.
7. Al-Jubouri, A. K. (2020). *Study of the effectiveness of alcoholic extract and nanopreparation of Dodonaea viscosa L leaves in some aspects of life and death events. For the major wax worm: Galleria mellonella L. (Lepidoptera Pyralidae)* ". M.S.C Thesis - College of Agriculture - University of Baghdad.126p.
8. Almediy, A. A.; Nenaah, G. E. & Shower, D. M. (2018). Facile synthesis of silver nanoparticles using harmala alkaloids and their insecticidal and growth inhibitory activities against the khapra beetle. *Journal of Pest Science*. 91: 727–737.
9. Al-Mashhadani, M. Mu. Ma. (2012). Study of the effect of powders of some plants, the pesticide Persquet and the fungus Beauveria bassiana on the life of the insect of the beetle Trogoderma granarium Everts (Coleoptera: Dermestidae). Master's thesis. Faculty of Education. Tikrit University. Iraq.110p.
10. Al-Qurashi, S. A.; Al-Saqqaf, A. I. & Mahyoub, J. A. (2015). *BioEffects of Silver Nanofine Particles Prepared from Thevetia neriifolia Yellow Oleander Extract Against Mosquito Larvae. Culex pipiens L.* College of Science, Medical University, Saudi Arabia. 27(2): 13 – 23.
11. Al-Shammari, H. A. (2015). *The effect of the predator (Diptera: Cecidomyiidae) Dicrodiplosis manihoti Harris and silver nanoparticles prepared by biological methods on some aspects of the life of citrus mealybugs (Hemiptera: pseudococcidae) Planococcus citri* Risso. Ph.D. Thesis. College of Agriculture. University of Baghdad, 225 p.
12. Al-Shujairi, A. K. R. (2018). *Evaluation of the efficacy of some nanopreparations and conventional in affecting a cotton insect Aphis gossypii Glover (Hemiptera:Aphididae) and the predator Coccinella septempunctata L. on different plant additives*. M.S.C Thesis. college of Agriculture. University of Baghdad.115 pages.
13. Alvi, A. M.; Iqbal, N.; Bashir, M.A.; Rehmani, M. I. A.; Ullah, Z.; Latif, A. & Saeed, Q. (2018). Efficacy of *Rhazya stricta* leaf and seed extracts against *Rhyzopertha dominica* and *Trogoderma granarium*. *Kuwait Journal of Science*. 45(3): 64 -71.
14. Athanassiou, C.G.; Phillips, T.W. & Wakil, W. (2019). Biology and control of the khapra beetle, *Trogoderma granarium*, a major quarantine threat to global food security". *Annual review of entomology*. 64: 131-148.
15. Chakravarthy, A. K.; Chandrashekharaiyah, S. B.; Kandakoor, B.; Dhanabala, K.; Gurunatha, K. and Ramesh, P. (2012). Bio efficacy of inorganic nanoparticles CdS. Nano - Ag and Nano - TiO₂ *Spodoptera litura* (Fabricius) (Lepidoptera: Noctuidae). *Current Biotica*. 6(3):271- 281.
16. Chen, H.; Wang, B.; Feng, W.; Du, W.; Ouyang, H.; Chai, Z. & Bi, X. (2014). Oral magnetite nanoparticles disturb the development of *Drosophila melanogaster* from oogenesis to adult emergence. *Nanotoxicology*. 5390:1-11.



17. Gulzar, M. U.; Threem, Z.; Muhammad, S. & Usama, I. M. (2020). Impacts of bio synthesized silver-nanoparticles (AgNO₃) and plant oils against *Trogoderma granarium*. *GSC Biological and Pharmaceutical Sciences*. 10 (03): 10-15.
18. Hadizadeh, I.; Peivastegan, B. & Kolahi, M. (2009). Antifungal activity of Nettle (*Urtica dioica* L.), Colocynth (*Citrullus colocynthis* L. Schrad), Oleander (*Nerium oleander* L.) and Konar (*Ziziphus spina-christi* L.) extract on plants pathogenic fungi. *Pakistan Journal of Biological Sciences*. 12(1): 58 – 63.
19. Khalique, U.; Farooq, M.U.; Ahmed, M.F. & Niaz, U. (2018). Khapra Beetle: A Review of Recent Control Methods. *Current Investment Agricultural*. Current Research. 5 (5):666-671.
20. Khan, S. A.; Ranjha, M. H.; Khan, A. A.; Sagheer, M.; Abbas, A. & Hassan, Z. (2019). Insecticidal efficacy of wild medicinal plants, *Datura alba* and *Calotropis procera*, against *Trogoderma granarium* (Everts) in wheat store grains. *Pakistan Journal of Zoology*. 51 (1) 289-294.
21. Komalamisra, N.; Trongtokit, Y.; Rongsriyam Y. & Apiwathnasorn C. (2005). Screening for larvicidal activity in some Thai plants against four mosquito vector species. *Southeast Asian Journal of Tropical Medicine and Public Health*. 36(6):1412-1422.
22. Magro, M.; Bramuzzo, S.; Baratella, D.; Ugolotti, J.; Zoppellaro, G.; Chemello, G.; Olivotto, I.; Ballarin, C.; Radaelli, G. & Arcaro, B. (2019). Self-assembly of chlorin-e6 on γ -Fe₂O₃ nanoparticles: Application for larvicidal activity against *Aedes aegypti*. *Journal Photochem. Photobiol. B. Biol.* 194: 21-31.
23. Media. (2007). *Russia suspends import of Indian plant products*. PTI, Moscow, posted: Wed, Jan 23, 4:26 PM IST.
24. Murugan, K.; Dinesh, D.; Nataraj, D.; Subramaniam, J.; Amuthavalli, P.; Madhavan, J.; Rajasekar, A.; Rajan, M.; Thiruppathi, K. P.; Kumar, S.; Higuchi, A.; Nicoletti, M. & Benelli, G. (2018). "Iron and iron oxide nanoparticles are highly toxic to *Culex quinquefasciatus* with little non target effects on larvivorous fishes". *Journal Environmental Science Polling Research*. 25(11): 10504-10514.
25. Mustafa, M. K. (2017). *Technology of the grain industry and its products*. Academic Library. Iraq
26. Najib, M. S. (2019). *Evaluation of the efficacy of nanocontrol of biological control against immature phases of the major wax moth. Galleria mellonella (L.) (Lepidoptera: Pyralidae)*. M.S.C Thesis. College of Education for Girls, University of Kufa. Iraq.
27. Nashwa, S.M. & Abo-Elyousr, K. A. M. (2012). Evaluation of various plants extract against the early blight disease of tomato plants under greenhouse and field condition. *Plant Protection, Science*. 48(2): 74 – 79.
28. United Nations (UN). (2022). *La population mondiale atteindra 8 milliards cette année malgré le ralentissement du taux de croissance*. *Population. Croissance démographique*. Bulletin.



29. Ovais, M.; Khalil, A.; Ayaz, M.; Ahmad, I. Nethi, S. K. & Mukherjee, S. (2018). Biosynthesis of Metal Nanoparticles via Microbial Enzymes: A Mechanistic Approach, *International Journal Molecular Sciences*. 19(12): 10- 40.
30. Rajendran, S. (2020). Insect Pest Management in Stored Products. *Outlooks on Pest Management*. 31(1): 24-35.
31. Raveen, R.; Kamakshi, K. T.; Deepa, M.; Arivoli, S. & Samuel, T. (2014). Larvicidal activity of *Nerium oleander* L. (Apocynaceae) flower extracts against *Culex quinquefasciatus* Say (Diptera: Culicidae). *International journal of mosquito Research*. 1(1): 38 – 42.
32. Reda, F. A.; Noha, A.; Mohammed, S. & Ibrahim, R. (2016). Toxicological Effects of Hematite Nanoparticles on the Common House Mosquito, *Culex pipiens* L. (Diptera: Culicidae). *Egyptian Academic Journal Biological Sciences*. 9(1): 57- 62.
33. Rouhani, M.; Samih, M. & Kalantari, A. S. (2008). Insecticidal effect of silica and silver nanoparticles on the cowpea seed beetle, *Callosobruchus maculatus* F. (Col.: Bruchidae). *Journal of Entomological Research*. 4(4): 297- 305.
34. Sahayaraj, K. (2017). Nano and Bio - nanoparticles for Insect Control. *Research Journal of Nanoscience and Nanotechnology*. 7(1): 1-9.
35. Sahi, S. N. (2019). *Study of the effect of ethyl alcohol extract and petroleum ether of Sesbania sesban L. leaf on the different roles of the capillary grain beetle (Trogoderma granarium (Everts) (Coleoptera: Dermestidae)*. M.S.C Thesis. Department of Plant Protection. college of Agricultural Engineering Sciences. University of Baghdad. 91p.
36. Suleiman, A. K. (2005). *Toxicity of some plant extracts Eucalyptus camaldulensis Dehnh. Eucalyptus, Swimmer Melia azedarach L. and Oleander. Nerium oleander L on the life of the rusty flour beetle insect Tribolium castaneum (Herbst) (Coleoptera: Tenebrionidae)*. M.S. C thesis. Faculty of Education. University of Baghdad. iraq.
37. Tawfeeq, A. T. (2014). Diluted concentrations of large (above one hundred nanometer) silver nanoparticles inhibited the growth of different types and origin of cancer cells". *Iraqi Journal of Cancer and Medical Genetics*. 7(1): 69-76.
38. Vecchio, G.; Galeone, A.; Malvindi, M. A.; Cingolani, R. & Pompa, P. P. (2013). Ranking the in vivo toxicity of nanomaterials in *Drosophila melanogaster*. *Journal Nanoparticle Research*. 15:1963.
39. Vega-Alvarez, S.; Herrera, A.; Rinaldi, C.; & Carrero-Martinez, F. A. (2014). Tissuespecific direct microtransfer of nanomaterials into *Drosophila* embryos as a versatile in vivo test bed for nanomaterial toxicity assessment. *Intrnational Journal Nanomedicine*. 9(1): 2031- 2041.
40. Wakil, W.; Kavallieratos, N.G.; Usman, M.; Gulzar, S. & El-Shafie, H. A. F. (2021). Detection of Phosphine Resistance in Field Populations of Four Key Stored-Grain Insect Pests in Pakistan. *Insects*. 12(4): 288 -299.



41. Yılmaz, A. & Koçak, E. (2022). Comparing bioassay and diagnostic molecular marker for phosphine resistance in Turkish populations of *Rhyzopertha dominica* (F., 1792) (Coleoptera: Bostrichidae). *Turkish Journal of Entomology*. 46(4): 431 – 440.
42. Zahir, A. A.; Bagavan, A.; Kamaraj, C.; Elango, G. & Rahuman, A. A. (2012). Efficacy of plant-mediated synthesized silver nanoparticles against *Sitophilus oryzae*. *Journal Biopest*. 5: 95 - 102.
43. Zhou, Y.; Rocha A.; Sanchez, C. J. & Liang, H. (2012). *Methods in Molecular Biology*, Walker, J. M. (Series Editor), *Nanoparticles in Biology and Medicine: Methods and Protocols*, Soloviev, M.(editior), Chapter 35 *Assessment of Toxicity of Nanoparticles Using_Insects as Biological Models*. Humana Pressa, Springer Science and Business Media,LLC, 233 Spring St.,New York. 906: 423- 434.